Fixation of Natural Furanoeremophilane by Diels-Alder Reaction

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Diels–Alder reaction of furanoeremophilan- 6β -ol (petasalbin) and its synthetic analogue, 4-hydroxy-4,5,6,7-tetra-hydrobenzofuran, was studied using *N*-ethyl- and *N*-phenylmaleimides as the dienophiles, affording corresponding adducts as mixtures of stereoisomers. The adduct of petasalbin and maleimide was also obtained when the latter compound was added to a crude extracted solution of *Petasites japonicus* var. *giganteus*. By using this method, it was estimated that the contents of unstable furanoeremophilan- 10β -ol in *L. cymbulifera* is more than 32% of the extract.

Furanoeremophilane compounds are found in nature in some plants of Compositae tribe Senecioneae, such as Ligularia, 1 Petasites, ² Farfugium, ³ and Syneilesis. ⁴ Although many compounds have been isolated in pure form and their structures have been determined, a problem is present in the handling of these compounds because some of them are unstable. For example, on the course of our investigation on the diversity in Ligularia species in the Hengduan Mountains area,⁵ we have found the presence of extremely unstable compound in L. tongolensis, but the compound has not been isolated.^{5a} Some unstable furanoeremophilanes decompose even in CDCl₃ solution, and therefore, many NMR data have been acquired in C₆D₆. 1c,e,3,4 The instability of these compounds are due to the presence of an electron-rich trisubstituted furan ring, which reacts easily with electron deficient reagents, such as proton. For the structure determination of these unstable compounds, the problem is to obtain the targeted molecules in pure form.

A possible solution to this problem is to "fix" the furan ring by conversion to some stable functional group. To this purpose, we planned to use a Diels–Alder reaction for the transformation of the furan ring into stable adduct, because furan is a good diene,⁶ and especially, the furan ring of natural furanoeremophilanes is electron-rich and therefore the Diels–Alder reaction is expected to proceed smoothly. A natural intramolecular Diels–Alder adduct of 1-acyloxyfuranoeremophilane derivative has been reported, indicating that the reaction proceeds without heating.⁷ Here, we report Diels–Alder reaction of natural petasalbin and synthetic analogue with *N*-phenyl- and *N*-ethyl-maleimide.

Results and Discussion

In the present study, two substrates, furanoeremophilan- 6β -ol (=ligularol or petasalbin, 1)^{8,9} and a synthetic analogue 2 were used. Compound 1 was isolated from the rhizome of the cultivated plant *Petasites japonicus* var. *giganteus* (Compositae), ¹⁰ which was collected in Nagano Prefecture, Japan. The synthetic analogue 2 was prepared by the Feist–Benary method ¹¹ from 1,3-cyclohexanedione and chloroacetaldehyde followed by LiAlH₄ reduction, as described previously (Chart 1). ⁸

Chart 1.

At first, the reaction of model compound 2 was examined with N-ethyl- and N-phenylmaleimide as the dienophiles (Scheme 1). The reaction was carried out in EtOH/H₂O 9:1 as the solvent at room temperature, which is a model condition of extraction of natural products, since the extracted solutions of natural products sometimes contain water and higher reaction temperature must be avoided for the treatment of unstable compounds. When 2 was treated with 3 molar amounts of Nethylmaleimide (3) under these conditions, the adduct was afforded in good yield as a mixture of three diastereomers. The isomers of the products were partly separated by silica-gel column chromatography giving a mixture of exo-adducts 4a and 4b (57% yield, ratio 7:3), and an isomer of endo-adduct 4c (41% yield). The fourth possible stereoisomer was not detected. The stereochemistry of the products was determined from ¹H NMR spectra. Namely, **4a** and **4b** were determined to be exo-adducts based on the J value between H_a and H_b $(J = 0 \,\mathrm{Hz})$, which indicated that $\mathrm{H_b}$ is endo to 7-oxabicyclo-[2.2.1]heptane ring system. The orientation of the hydroxy group was axial for 4a and equatorial for 4b, respectively, judged from the coupling constants of the proton attached to the hydroxy-bearing carbon. However, since two conformers were possible for the cyclohexane ring, it was difficult to determine the relative stereochemistry of 4a and 4b. Compound 4c was judged to be endo-adduct based on a relatively large J value (5.5 Hz) between H_a and H_b .

The reaction of **2** with *N*-phenylmaleimide (**5**) was also carried out similarly. Formation of adduct **6** was observed on TLC; however, in contrast to **4a–4c**, it was difficult to obtain each products in pure form, because a retro-Diels–Alder reaction occurred during the isolation process. Some other dieno-

Scheme 1. Reaction conditions: in EtOH/H₂O (9:1), r.t., 1 day.

Scheme 2. Reagents and conditions: i) benzoyl chloride, pyridine, CH₂Cl₂, r.t., 2 h. ii) see Table 1.

philes, such as benzoquinone, ethyl acrylate, diethyl fumarate, maleic anhydride, and diethyl acetylenedicarboxylate, were tested other than maleimide. However, none of them afforded the corresponding adducts in better yields than *N*-ethylmaleimide.

In order to show the utility of N-phenylmaleimide, the reaction was further studied using benzoate 7 as the substrate, which was prepared by benzoylation of 2 (Scheme 2). The

Table 1. Reaction of 7 and 5 under Various Conditions^{a)}

Entry	Equiv. of 5	Time/d	Yield/%
1	1	1	55
2	1	3	78
3	2	1	69
4	2	3	80
5	3	1	84
6	3	3	80

a) All reactions were carried out in $EtOH/H_2O$ (9:1) with 0.1 mol L^{-1} concentration of diene at room temperature.

adducts were obtained as a mixture of two stereoisomers, and the each diastereomer was stable enough for the isolation procedure to obtain **8a** (51%) and **8b** (14%) after purification on silica gel. Both products were determined to be exo-adducts as described above.

Optimization of the reaction condition was made for the reaction of 7 to 8. The results are shown in Table 1. The use of a stoichiometric amount of maleimide 5 was found to be enough if the reaction was carried out for a longer time (Entry 2). However, in the present study, we chose to use 3 equivalent of maleimide (Entry 5) in order to use natural product more effectively.

Following the succession with the model compound, the Diels-Alder reaction of natural furanoeremophilane was studied. When compound 1 was treated with 3 in EtOH/ H₂O (9:1) at room temperature, a pair of diastereomers 9a and 9b were obtained in 39 and 16% yields, respectively (Scheme 3). In contrast to the case of model compound 2, the adducts with N-phenylmaleimide (5) were obtained in better yield (74%) without the occurrence of the retro-Diels-Alder reaction. The ratio of the two diastereomers 10a and **10b** was 4:1. In both cases, stereoisomers of the two products were obtained, and each isomer could be isolated by silicagel column chromatography. The stereochemistry of the all four products were determined to be exo-adducts based on ¹HNMR spectra as described for the model compounds. The major isomer (9a/10a) and the minor isomer (9b/10b) were determined to be the β -exo and the α -exo adducts, respectively, based on the presence or absence of long-range coupling between olefinic methyl group and the proton attached to the hydroxy-bearing carbon. 12 These protons were almost planar with the C=C double bond for 9a/10a (J = 0 Hz) but not for **9b/10b** ($J = 2.2 \,\text{Hz}$).

The stereoselective formation of the exo-isomers can be rationalized as follows. Although four approaching directions of dienophile 3 (or 5) to 1 are possible, as shown in Scheme 4, β -endo and α -endo approaches are unfavorable because of steric congestion. Namely, the R group on the maleimide comes close to the C-6 substituent in the β -endo approach. In the α -endo approach, maleimide molecule must come inside the "umbrella" shape of the cis-decaline system of the furanoere-mophilane. Among two exo-approaches, β -exo is more favorable than α -exo because steric interaction with decaline ring is minimum. The conformation of furanoeremophilane illustrated in Scheme 4 is "steroidal," and there is another conformer, "non-steroidal." The story described here must be the same for the non-steroidal conformation, except that steric interac-

Scheme 3. Reaction conditions: in EtOH/H₂O (9:1), r.t., 1 day.

10b

10a

tion between C-5 methyl and dienophile must be considered for the β -endo approach. For the reaction of the model compound, related explanation can be applied for the formation of **4a**, **4b**, **8a**, and **8b**. The formation of α -endo product (**4c**) can be explained by the lack of interaction with the fused cyclohexane ring.

The present Diels-Alder reaction was applied to transform natural furanoeremophilane into stable adduct before the isolation procedure. At first, the utility of the present "fixation" method was estimated by comparison with the standard isolation using the same extracted solution. When 12 cm³ of AcOEt extracted solution of half-dried roots of P. japonicus var. giganteus was treated with N-ethylmaleimide (3), 6.6 mg of the adducts **9a** and **9b** (correspond to 4.3 mg of **1**) were isolated. The same treatment with N-phenylmaleimide (5) afforded 8.4 mg (correspond to 4.8 mg of 1) of 10a and 10b. In contrast, only 1.5 mg of 1 could be isolated by direct column chromatography from the same amount of the extracted AcOEt solution, although acidic conditions were avoided by using neutral silica gel. In order to reduce handling time after harvesting, the method was also applied to "fix" compound 1 in fresh root. When 3 was added to a crude extracted ethanol solution of the fresh root, a spot of the adduct 9 immediately appeared on TLC; however, it was difficult to judge the disappearance of the spot of 1. From 12 g of the fresh root, 9a (54.5 mg) and **9b** (19.4 mg) were afforded as crystals.

Me
$$\beta$$
-exo

R

9a

(or 10a)

Begin for 10b)

α-exo

Scheme 4. Stereochemistry of the reaction of 1 and 3 (or 5).

We have recently studied chemical constituents of L. cymbulifera, and four furanoeremophilanes have been isolated from the plant. 5a Among them, furanoeremophilan- 10β -ol (11) has been detected as the major component on TLC of the crude extracted solution, but the compound is too labile. From 1.5 g of the extract, only 176.3 mg (12%) of the compound has been obtained. Although we have not stated in the report, the compound decomposes when the solvent is evaporated after purification. Ehrlich's reaction of 11 was faster than the other furanoeremophilanes, indicating that the furan ring in 11 is highly reactive. Then, the present Diels-Alder method was applied to "fix" the unstable molecule. When N-ethylmaleimide was added to a crude extracted solution of L. cymbulifera, the reaction immediately occurred, and the spot of 11 on TLC disappeared. The other Ehrlich-positive spot did not disappear in a short reaction time, indicating that only 11 had been reacted. Form $10\,\mathrm{cm}^3$ of the extracted solution, the adduct (119.1 mg) was obtained as a mixture of two diastereomers 12a and 12b (Scheme 5). Since 2 cm³ of the extracted solution gave 48.7 mg of the residual oil, the result implies that the content of 11 is more than 32% of the extract. Interestingly, minor isomer 12b was not α -exo-adduct, but β -endo-adduct, which was determined from the J value

Scheme 5. Diels–Alder reaction of furanoeremophilan- 10β ol in *L. cymbulifera*.

Scheme 6. Stereochemistry of the formation of 12b.

(5.5 Hz) between the proton attached to the ethereal oxygenbearing carbon and the adjacent proton, as in the case of 4c. This unexpected stereoselectivity can be rationalized by the formation of hydrogen-bond between hydroxy group at C-10 and the carbonyl oxygen of the maleimide for the β -endo approach (Scheme 6).

In conclusion, the Diels-Alder reaction of natural furanoeremophilane compound with *N*-ethyl- and *N*-phenyl-maleimide was found to proceed smoothly at room temperature; however, *N*-ethylmaleimide was found to be better for the model compound. Although the occurrence of retro-Diels-Alder reaction in some case indicates limited usefulness, the method was shown to be useful for fixation of unstable natural furanoeremophilanes.

Experimental

General Procedure. Melting points were measured on a Laboratory Devices Mel-Temp apparatus. IR spectra were recorded on a Jasco FT/IR-230 spectrometer. Both 1 H and 13 C NMR spectra were measured on a Jeol GSX-400 (400 MHz for 1 H; 100 MHz for 13 C) spectrometer in CDCl₃ as the solvent, unless otherwise noted. Chemical shifts were recorded on the δ scale (ppm) with tetramethylsilane as an internal standard. For 13 C NMR, the signal of the solvent (CDCl₃: 77.0 ppm) was used as the reference. Both low-resolution mass spectra (MS) and high-resolution mass spectra (HRMS) were obtained on a Jeol SX-102A, CMATE II, JMS AX-500, or Shimadzu GCMS-QP5050 mass spectrometer. Analytical TLC was done on precoated TLC plates (Kieselgel 60 F254, layer thickness 0.2 mm).

Wakogel C-200 was used for column chromatography unless otherwise noted.

Materials. Compound **1** was isolated from roots of *Petasites japonicus* var. *giganteus* (Compositae) collected in Nagano Prefecture, Japan. See our previous report⁸ for the isolation procedure. Compound **2** was prepared via the known Feist–Benary procedure. ^{8,11}

4-Benzoyloxy-4,5,6,7-tetrahydrobenzofuran (7). In a 50 cm³ round-bottomed flask attached with CaCl2 drying tube, a stirred solution of 2 (1.90 g, 13.8 mmol) in CH₂Cl₂ (50 cm³) was prepared. To this was added pyridine (5.8 cm³) and benzoyl chloride (5.00 cm³, 61.8 mmol) successively, and the stirring was continued at room temperature for 2 h. Water (40 cm³) was added, and the mixture was extracted with CH2Cl2 and dried over anhydrous MgSO₄. Evaporation of the solvent, followed by silicagel column chromatography using hexane/AcOEt (99:1) as eluent. afforded 7 (3.19 g. 96%) as an oil: IR (neat) 2949, 1714. 1271, 1109, and 712 cm⁻¹; ¹H NMR (C_6D_6) δ 1.30–1.40 (1H, m), 1.49-1.58 (1H, m), 1.69-1.84 (2H, m), 2.12-2.21 (1H, m), 2.33-2.41 (1H, m), 6.06 (1H, t, J = 3.8 Hz), 6.44 (1H, d, J = $1.5\,Hz),\ 6.95-7.13\ (4H,\ m),\ and\ 8.14-8.18\ (2H,\ m);\ ^{13}C\,NMR$ (CDCl₃) δ 19.01, 22.76, 28.97, 66.71, 110.08, 116.42, 128.09 (2C), 129.41 (2C), 130.46, 132.62, 140.77, 153.84, and 166.09; MS (EI) m/z 137 (M⁺ – PhCO), 120 (base), 105, 91, and 77; HRMS (EI) Found: m/z 242.0943 (M⁺); Calcd for $C_{15}H_{14}O_3$: M, 242.0943.

Diels–Alder Reaction of 2 and 3. Compound **2** (137.1 mg, 0.993 mmol) was dissolved in EtOH/ H_2O (10 cm³, ratio 9:1), and to this was added a solution of *N*-ethylmaleimide (**3**, 378.9 mg, 3.03 mmol) in EtOH/ H_2O of the same ratio (10 cm³) at room temperature with stirring, which was continued for 1 day. Without extraction, water was eliminated from the mixture by adding Na₂SO₄, followed by filtration. Evaporation of the solvent afforded an oily residue, which was chromatographed on silica gel (35 g) using hexane/AcOEt (8:2 to 2:8) as the eluent to afford a mixture of **4a** and **4b** (148.5 mg, 57%) and **4c** (106.3 mg, 41%).

A mixture of **4a** and **4b**: Mp 138–140 °C; IR (KBr) 3377 (OH), 1695 (C=O), 1408, 1344, 1232, and 1142 cm⁻¹; ¹H NMR δ 1.14 (3H, t, $J = 7.2 \,\text{Hz}$, NCH₂CH₃), 1.40–2.21 (6H, m), 2.51–2.63 (1H, m), 2.72 (1H \times 3/10, d, J = 6.5 Hz, H_c of **4b**), 2.98 (1H \times 7/10, d, J = 6.5 Hz, H_c of **4a**), 3.00 (1H × 3/10, d, J = 6.5 Hz, H_b of **4b**), 3.11 (1H × 7/10, d, J = 6.5 Hz, H_b of **4a**), 3.51 (2H, q, J = 7.2 Hz, NC H_2 CH₃), 4.20 (1H × 3/10, ddd, J = 2.5, 5.5, 11.6 Hz, CHOH of **4b**), 4.71 (1H \times 7/10, t, J = 2.5 Hz, CHOH of 4a), 5.13 (1H \times 7/10, d, J = 1.6 Hz, H_a of 4a), 5.14 (1H \times 3/10, d, J = 1.6 Hz, H_a of **4b**), 6.30 (1H × 3/10, br s, C=CH of **4b**), and 6.32 (1H \times 7/10, s, C=CH of **4a**); ¹³C NMR assigned for **4a**: δ 12.91, 16.50, 26.21, 32.45, 33.74, 47.63, 50.93, 64.26, 79.59, 87.80, 131.36, 149.99, 174.87, and 176.18; assigned for **4b**: δ 12.94, 20.41, 25.74, 33.81, 34.73, 46.76, 51.80, 67.98, 79.88, 89.58, 127.47, 153.30, 174.70, and 175.91; MS (CI) m/z 264 (M⁺ + H), 246, 166, 138, and 121 (base); HRMS (CI) Found: m/z 264.1235 (M⁺ + H); Calcd for C₁₄H₁₈NO₄: M, 264.1236.

4c: Mp 111–112 °C; IR (KBr) 3438 (OH), 1697 (C=O), 1402, 1342, 1225, 1066, and 895 cm⁻¹; ¹H NMR δ 1.05 (3H, t, J = 7.2 Hz, NCH₂CH₃), 1.34–2.43 (7H, m), 3.20 (1H, d, J = 7.4 Hz, H_c), 3.39 (2H, q, J = 7.2 Hz, NCH₂CH₃), 3.68 (1H, dd, J = 5.5, 7.4 Hz, H_b), 3.99 (1H, br, $W_{1/2}$ = 23 Hz, CHOH), 5.17 (1H, dd, J = 1.6, 5.5 Hz, H_a), and 6.19 (1H, br s, C=CH); ¹³C NMR δ 12.59, 19.76, 30.66, 33.33, 34.70, 50.02, 50.98, 68.27, 78.01,

89.77, 125.32, 151.38, 174.90, and 174.97; MS (CI) m/z 263 (M⁺), 246, 166, 118, 126, and 121 (base); HRMS (CI) Found: m/z 264.1237 (M⁺ + H); Calcd for C₁₄H₁₈NO₄: M, 264.1236.

Diels–Alder Reaction of 7 and 5. By the same procedure described above, compound **7** (50.5 mg, 0.209 mmol) and **5** (36.2 mg, 0.209 mmol) in EtOH/ $\rm H_2O$ afforded **8a** (44.5 mg, 51%) and **8b** (12.4 mg, 14%) after silica-gel chromatography using hexane/AcOEt (2:1) as the eluent.

8a: An oil; IR (neat) 1712 (C=O), 1385, 1265, 1192, and 737 cm⁻¹; ¹H NMR δ 1.74–2.23 (5H, m), 2.73 (1H, br d, J = 13 Hz), 2.97 (1H, d, J = 6.5 Hz, CHC=O), 3.07 (1H, d, J = 6.5 Hz, CHC=O), 5.23 (1H, d, J = 1.6 Hz, CH-O), 5.97 (1H, br s, CHOBz), 6.55 (1H, br s, C=CH), 7.21–7.56 and 7.95–7.99 (10H, m, Ph × 2); ¹³C NMR δ 17.56, 25.98, 29.97, 47.49, 50.43, 66.74, 80.20, 88.11, 126.44 (2C), 128.56 (2C), 128.60, 129.00 (2C), 129.36 (2C), 129.85, 131.61, 133.26, 135.23, 145.91, 165.02, 173.79, and 174.88; MS (FAB) m/z 438 (M⁺ + Na), 416 (M⁺ + H), 413, 329, 307, 154 (base), and 136; HRMS (FAB) Found: m/z 416.1485 (M⁺ + H); Calcd for C₂₅H₂₂NO₅: M, 416.1499.

8b: Mp 169–170 °C; IR (neat) 1712 (C=O), 1385, 1265, and 737 cm⁻¹; ¹H NMR δ 1.67–1.95 (3H, m), 2.08–2.16 (1H, m), 2.29–2.36 (1H, m), 2.69 (1H, br d, $J=13\,\mathrm{Hz}$), 3.01 (1H, d, $J=6.5\,\mathrm{Hz}$, CHC=O), 3.19 (1H, d, $J=6.5\,\mathrm{Hz}$, CHC=O), 5.28 (1H, d, $J=1.5\,\mathrm{Hz}$, CH=O), 5.65 (1H, ddd, J=2.6, 5.9, 11.2 Hz, CHOBz), 6.26 (1H, br s, C=CH), 7.25–7.64 and 8.05–8.09 (10H, m, Ph × 2); ¹³C NMR δ 20.33, 25.76, 30.76, 46.93, 51.61, 69.25, 80.48, 90.19, 126.51 (2C), 128.51 (2C), 128.58, 128.72, 129.09 (2C), 129.57, 129.71 (2C), 131.65, 133.41, 148.96, 165.72, 173.72, and 175.04; MS (FAB) m/z 438 (M⁺ + Na), 416 (M⁺ + H), 413, 391, 294, 154 (base), and 121; HRMS (FAB) Found: m/z 416.1482 (M⁺ + H); Calcd for C₂₅H₂₂NO₅: M, 416.1499.

Diels–Alder Reaction of Furanoeremophilan-6 β -ol (1). In a 10 cm³ round-bottomed flask, a solution of compound 1 (13.1 mg, 0.056 mmol) in EtOH/H₂O (0.3 cm³, ratio 9:1) was prepared, and to this was added a solution of 5 (30.3 mg, 0.175 mmol) in the same solvent (0.3 cm³) with stirring at room temperature. After being stirred for 24 h, water was removed by adding anhydrous Na₂SO₄ followed by filtration. The solvent EtOH was evaporated, and the resultant residue was chromatographed on silica gel (3 g) using hexane/AcOEt (4:1 to 0:1) as the eluent to afford a mixture of 10a and 10b (16.8 mg, 74%). Similarly, the reaction of 1 (12.5 mg, 0.0533 mmol) and 3 (23.0 mg, 0.184 mmol) in EtOH/H₂O (0.5 cm³) afforded 9a (7.7 mg, 39%) and 9b (3.2 mg, 16%) after silica-gel (2 g) column chromatography using hexane/AcOEt (7:3 to 4:6) as eluent.

9a: Mp 70–72 °C; IR (KBr) 3448, 2927, 1693, 1406, and 999 cm⁻¹; 1 H NMR δ 0.69 (3H, d, J=6.5 Hz, Me), 1.08 (3H, s, Me), 1.14 (3H, t, J=7.2 Hz, NCH₂CH₃), 1.23–1.94 (8H, m), 1.86 (3H, s, C=CMe), 2.22–2.29 (3H, m), 2.91 (1H, d, J=6.5 Hz, CHC=O), 3.13 (1H, d, J=6.5 Hz, CHC=O), 3.51 (2H, q, J=7.2 Hz, NCH₂CH₃), 4.39 (1H, br s, CHOH), and 4.89 (1H, s, CHOC); 13 C NMR δ 9.92, 12.97, 15.76, 16.53, 19.44, 25.50, 25.68, 29.50, 30.29, 32.69, 33.71, 40.09, 49.83, 50.29, 68.77, 83.39, 88.79, 140.20, 141.67, 175.37, and 176.60; MS (CI) m/z 360 (M⁺ + H), 342, 234, 217 (base), 126, and 124; HRMS (CI) Found: m/z 360.2172 (M⁺ + H); Calcd for C₂₁H₃₀NO₄: M, 360.2176.

9b: Mp 151–152 °C; IR (KBr) 3448, 2925, 1697, and 1400 cm⁻¹; ¹H NMR δ 0.91 (3H, s, Me), 1.11 (3H, d, J = 7.1 Hz, Me), 1.13 (3H, t, J = 7.2 Hz, NCH₂CH₃), 1.19–2.10 (10H, m),

2.01 (3H, d, $J=2.2\,\mathrm{Hz}$, C=CMe), 2.25 (1H, dd, J=2.2, 13.8 Hz), 2.75 (1H, d, $J=6.5\,\mathrm{Hz}$, CHC=O), 2.92 (1H, d, $J=6.5\,\mathrm{Hz}$, CHC=O), 3.50 (2H, q, $J=7.2\,\mathrm{Hz}$, NC $H_2\mathrm{CH}_3$), 4.78 (1H, br, $W_{1/2}=14\,\mathrm{Hz}$, CHOH), and 4.78 (1H, s, CHOC); $^{13}\mathrm{C}\,\mathrm{NMR}$ & 11.05, 12.97, 14.86, 18.94, 20.78, 27.05, 28.56, 29.32, 31.89, 33.78, 37.24, 41.50, 49.98, 51.22, 70.34, 83.88, 89.77, 140.21, 142.21, 174.98, and 176.33; MS (CI) m/z 360 (M⁺ + H), 342, 235, 217, 124 (base); Analysis Found: C, 70.24; H, 8.44; N, 3.61%; Calcd for C₂₁H₂₉NO₄: C, 70.17; H, 8.13; N, 3.90%.

10a: Mp 163–164 °C; IR (KBr) 3498, 2925, 1703, 1389, and 1190 cm⁻¹; ¹H NMR δ 0.70 (3H, d, J = 6.8 Hz, Me), 1.09 (3H, s, Me), 1.24–2.34 (11H, m), 1.90 (3H, s, C=CMe), 3.08 (1H, d, J = 6.5 Hz, CHC=O), 3.31 (1H, d, J = 6.5 Hz, CHC=O), 4.42 (1H, s, CHOH), 5.02 (1H, s, CHOC), and 7.26–7.48 (5H, m, Ph); ¹³C NMR δ 10.04, 15.80, 16.56, 19.42, 25.47, 25.73, 29.50, 30.28, 32.67, 40.09, 49.92, 50.36, 68.80, 83.87, 89.29, 126.58 (2C), 128.59, 129.06 (2C), 131.80, 141.33, 141.89, 174.56, and 175.91; MS (FAB) m/z 430 (M⁺ + Na), 413, 408 (M⁺ + H), 390, 217 (base), and 124; HRMS (FAB) Found: m/z 408.2155 (M⁺ + H); Calcd for C₂₅H₃₀NO₄; M, 408.2176.

10b: Mp 111–113 °C; IR (KBr) 3450, 2929, 1705, 1387, and 1190 cm⁻¹; ¹H NMR δ 0.93 (3H, s, Me), 1.02 (3H, d, J = 7.2 Hz, Me), 1.18–2.33 (11H, m), 2.05 (3H, d, J = 2.2 Hz, C=CMe), 2.92 (1H, d, J = 6.5 Hz, CHC=O), 3.10 (1H, d, J = 6.5 Hz, CHC=O), 4.82 (1H, br dq, J = 8.7, 1.9 Hz, CHOH), 4.92 (1H, s, CHOC), and 7.24–7.48 (5H, m, Ph); ¹³C NMR δ 11.15, 14.87, 18.98, 20.76, 27.11, 28.56, 29.28, 31.89, 32.23, 41.51, 50.04, 51.34, 70.37, 84.36, 90.27, 126.53 (2C), 128.65, 129.09 (2C), 131.72, 140.43, 142.41, 174.17, and 175.64; MS (FAB) m/z 430 (M⁺ + Na), 413, 408 (M⁺ + H), 390, 329, 307, 154 (base), and 136; HRMS (FAB) Found: m/z 408.2151 (M⁺ + H); Calcd for C₂₅H₃₀NO₄: M, 408.2176.

Diels-Alder Reaction of Unpurified Extract of Petasites japonicus var. giganteus. Comparison of the Diels-Alder Method and the Conventional Isolation: Half-dried roots (569 g) were extracted with AcOEt at room temperature as usual. The extracted solution (36 cm³) was obtained by decantation and was divided into three portions. To the first 12 cm³ portion was added a solution of N-ethylmaleimide (126.8 mg) in AcOEt (5 cm⁵), and the mixture was stirred at room temperature for 1 week. Evaporation of the solvent followed by silica-gel (2 g) column chromatography using hexane/AcOEt (9:1) as the eluent afforded a mixture of **9a** and **9b** (6.6 mg). The second 12 cm³ portion of the extracted solution was similarly treated with a solution of N-phenylmaleimide (175.7 mg) in AcOEt (5 cm³) giving 10a and 10b (8.4 mg). The third 12 cm³ portion was evaporated to an oily residue (52.4 mg), which was chromatographed on neutral silica gel (Silica Gel 60 N) (5 g) using hexane/AcOEt (9:1) as the eluent to afford 1 (1.5 mg).

Application to the Extraction of Fresh Root: Fresh roots (12 g) were extracted with ethanol (13 cm^3) for 1 week at room temperature. The plant material was filtered off, and to the filtrate was added *N*-ethylmaleimide (108.7 mg). The reaction mixture was stirred at room temperature for 1 week, followed by evaporation of the solvent. The resultant residue was chromatographed as described above to afford $\mathbf{9a}$ (54.5 mg) and $\mathbf{9b}$ (19.4 mg).

Diels–Alder Reaction of Unpurified Extract of Ligularia cymbulifera. Dried roots, collected near Zhongdian city, Yunnan, China, were extracted with ethyl acetate for 1 week at room temperature. An extracted solution $(10 \,\mathrm{cm}^3)$ was obtained by decantation, and a solution of N-ethylmaleimide $(2.8 \,\mathrm{g})$ in AcOEt $(4 \,\mathrm{cm}^3)$ was added. After keeping the mixture for 30 min at room

temperature, the solvent was evaporated off, and the residual oil was separated by silica-gel (8 g) column chromatography using hexane—AcOEt (9:1 to 3:1) as the eluent to afford a mixture of **12a** and **12b** (119.1 mg, ratio 3:2). The following spectral data were obtained after further separation.

At the same time, in order to calculate the content of 11, the extracted solution of L. cymbulifera (2 cm^3) was simply evaporated to yield an oily residue (48.7 mg).

12a: Mp 138–139 °C; IR (KBr) 3517 and 1682 cm⁻¹; ¹H NMR δ 0.77 (3H, d, J = 6.5 Hz, Me), 0.97 (3H, s, Me), 1.15 (3H, t, J = 7.1 Hz, NCH₂CH₃), 1.25–1.97 (8H, m), 1.74 (3H, d, J = 2.5 Hz, C=CMe), 2.09 (1H, d, J = 14.6 Hz), 2.10 (1H, dq, J = 16.6, 2.5 Hz), 2.33 (1H, d, J = 16.6 Hz), 2.63 (1H, d, J = 14.6 Hz), 2.86 (1H, d, J = 6.4 Hz, CHC=O), 2.90 (1H, d, J = 6.4 Hz, CHC=O), 3.51 (2H, q, J = 7.1 Hz, NCH₂CH₃), and 4.88 (1H, s, CHOC); ¹³C NMR δ 9.87, 12.98, 14.35, 15.71, 22.61, 28.62, 29.57, 33.33, 33.45, 33.72, 36.50, 41.36, 49.82, 50.93, 74.52, 80.07, 88.63, 135.89, 139.60, 176.30, and 176.62; MS (CI) m/z 360 (M⁺ + H), 342, 291, 235, 217 (base), 126, and 89; HRMS (CI) Found: m/z 360.2172 (M⁺ + H); Calcd for C₂₁H₃₀NO₄: M, 360.2176.

12b: Mp 129–131 °C; IR (KBr) 3479 and 1682 cm⁻¹; ¹H NMR δ 0.72 (3H, d, J = 6.5 Hz, Me), 0.97 (3H, s, Me), 1.02 (3H, t, J = 7.1 Hz, NCH₂CH₃), 1.11–1.90 (8H, m), 1.69 (3H, s, C=CMe), 2.14 (1H, d, J = 15.4 Hz), 2.30 (1H, br d, J = 16.5 Hz), 2.87 (1H, d, J = 15.4 Hz), 3.27 (1H, d, J = 7.3 Hz, CHC=O), 3.41 (2H, q, J = 7.1 Hz, NCH₂CH₃), 3.69 (1H, dd, J = 5.5, 7.3 Hz, CHC=O), 4.53 (1H, br, OH), and 4.95 (1H, d, J = 5.5 Hz, CHOC); ¹³C NMR δ 11.09, 13.13, 14.58, 15.60, 22.49, 29.34, 29.70, 33.48, 33.53, 33.88, 38.56, 42.89, 49.55, 52.49, 73.87, 81.62, 88.79, 134.93, 137.23, 174.31, and 177.37; MS (CI) m/z 360 (M⁺ + H), 342, 291, 235, 217 (base), and 126; HRMS (CI) Found: m/z 360.2184 (M⁺ + H); Calcd for C₂₁H₃₀NO₄: M, 360.2176.

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